

Mechanisms of quinolone resistance in *Escherichia coli* isolated from companion animals, pet-owners, and non-pet-owners

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The present study investigated the prevalence and mechanisms of fluoroquinolone (FQ)/quinolone (Q) resistance in *Escherichia (E.) coli* isolates from companion animals, pet-owners, and non-pet-owners. A total of 63 *E. coli* isolates were collected from 104 anal swab samples, and 27 nalidixic acid (NA)-resistant isolates were identified. Of those, 10 showed ciprofloxacin (CIP) resistance. A plasmid-mediated Q resistance gene was detected in one isolate. Increased efflux pump activity, as measured by organic solvent tolerance assay, was detected in 18 NA-resistant isolates (66.7%), but was not correlated with an increase in minimum inhibitory concentration (MIC). Target gene mutations in Q resistance-determining regions (QRDRs) were the main cause of (FQ)Q resistance in *E. coli*. Point mutations in QRDRs were detected in all NA-resistant isolates, and the number of mutations was strongly correlated with increased MIC ($R = 0.878$ for NA and 0.954 for CIP). All CIP-resistant isolates ($n = 10$) had double mutations in the *gyrA* gene, with additional mutations in *parC* and *parE*. Interestingly, (FQ)Q resistance mechanisms in isolates from companion animals were the same as those in humans. Therefore, prudent use of (FQ)Q in veterinary medicine is warranted to prevent the dissemination of (FQ)Q-resistant bacteria from animals to humans.

Keywords: *Escherichia coli*, antimicrobial drug resistance, fluoroquinolones, pets, quinolones

Introduction

Quinolone (Q) and fluoroquinolone (FQ) are broad-spectrum synthetic antimicrobials used to treat bacterial infections in humans and animals [23,27]. Since they are very potent antimicrobial agents against Gram-negative bacteria, including *Escherichia (E.) coli*, these agents have been widely used to treat a range of infections in human and veterinary medicine. Consequently, (FQ)Q resistance has markedly increased worldwide, posing a significant threat to the health of animals and humans [23,27].

Three major mechanisms of (FQ)Q resistance have been reported: (1) mutations in genes encoding DNA gyrase (*gyrA* and *gyrB*) and topoisomerase IV (*parC* and *parE*) that are associated with quinolone resistance-determining regions (QRDRs); (2) the presence of plasmid-mediated Q resistance (PMQR) genes; and (3) reduced accumulation of drugs or

chemicals due to active efflux pump activity [17]. PMQR genes include members of the *qnr* gene family (*qnrA*, *qnrB*, and *qnrS*) as well as genes encoding FQ-modifying enzyme [*aac-(6)-Ib-cr*] and the efflux pump (*qepA*) [40]. AcrAB-TolC overexpression is a major resistance mechanism against (FQ)Q that is associated with increased efflux pump activity and contributes to multi-drug resistance (MDR) in *E. coli* [28]. AcrAB-TolC has three components: a transporter of the resistance-nodulation-division family (AcrB), a periplasmic accessory protein (AcrA), and an outer membrane protein (TolC) [28].

E. coli are usually commensal bacteria in humans and animals. They are also considered major causative agents of bacterial infections. *E. coli* easily acquire antimicrobial resistance by genetic mutation and horizontal gene transfer [3]. The transmission of antimicrobial-resistant *E. coli* from animals to humans has been demonstrated [30]. Several studies have investigated multi-factorial (FQ)Q resistance mechanisms in *E.*

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coli isolated from humans and food-producing animals [19]. However, few studies have examined the prevalence and the resistance mechanisms of (FQ)Q-resistant *E. coli* from companion animals [13], and most of these have been limited to elucidating one or two of the above-mentioned (FQ)Q resistance mechanisms. Moreover, there have been no studies investigating the distribution of (FQ)Q-resistant *E. coli* in pets and pet-owners. Clarifying how (FQ)Q resistance develops and distributes in companion animals and the humans that they contact is important for understanding (FQ)Q resistance trends in veterinary medicine. To this end, the present study examined the frequency of nalidixic acid (NA)-resistant *E. coli* isolated from companion animals and their owners and investigated the three basic mechanisms of (FQ)Q resistance in these isolates relative to those obtained from non-pet-owners.

Materials and Methods

Sampling

Sampling was carried out with informed consent from owners of companion animals and other human subjects. A total of 104 anal swab samples were collected from four local veterinary clinics, one veterinary teaching hospital, and one local university in Seoul, Korea between April 2010 and November 2012. The sampling procedures that were used have been previously described [14]. Swab samples were obtained from 49 dogs, four cats, 14 dog owners, three cat owners and 34 non-pet-owners (Table 1). People living with and without pets at the time of sampling were designated as pet-owners and non-pet-owners, respectively. Owners were selected from among visitors of four local veterinary clinics and a veterinary teaching hospital; non-pet-owners were selected from among freshman students at a university. All protocols and procedures were approved by the institutional review board at the Seoul National University (IRB No. 1208/001-004).

Isolation of NA-resistant *E. coli* from swab samples

E. coli isolation and confirmation was carried out as previously described [6]. After isolating and identifying *E. coli* from swab

samples, 30- μ g NA antimicrobial disks (BD Biosciences, USA) were used to select NA-resistant isolates according to Clinical and Laboratory Standards Institute (CLSI) standards [7].

Antimicrobial resistance profiling of NA-resistant *E. coli* isolates

The susceptibility of 27 NA-resistant isolates to other antimicrobials was characterized by using the following antimicrobial disks (BD Biosciences, USA): ampicillin (AM; 10 μ g), amoxicillin/clavulanic acid (AMC; 20/10 μ g), ceftazidime (CAZ; 30 μ g), cefotetan (CTT; 30 μ g), imipenem (10 μ g), gentamicin (10 μ g), tetracycline (30 μ g), ciprofloxacin (CIP; 5 μ g), sulfamethoxazole/trimethoprim (1.25/23.75 μ g), chloramphenicol (C; 30 μ g), aztreonam (ATM; 30 μ g), ceftriaxone (CRO; 30 μ g), and cefotaxime (CTX; 30 μ g). Susceptibility or resistance to antimicrobials was determined according to CLSI standards [7]. *E. coli* ATCC 25922 was used as a reference strain (American Type Culture Collection, USA). MDR isolates were defined as isolates showing resistance to more than three different classes of antimicrobials [21].

NA-resistant *E. coli* isolates showing inhibition zone diameters of ≤ 25 mm against CRO were selected for use in the confirmation test for extended spectrum β -lactamase (ESBL) production [7]. Isolates were identified as ESBL-producing *E. coli* by applying the disk diffusion method using CAZ, CAZ/clavulanic acid (CL) (30/10 μ g), CTX, and CTX/CL (30/10 μ g); those isolates showing resistance to CAZ and/or CTX in combination with an increase in inhibition zone diameter of ≥ 5 mm for CAZ/CL and/or CTX/CL were defined as ESBL-producing *E. coli* [33]. In addition, the presence of the *bla*_{CTX-M} gene in ESBL-producing isolates was determined by performing polymerase chain reaction (PCR) with CTX-M universal primers [22].

Determination of minimum inhibitory concentrations (MICs) of NA and CIP

The MICs of NA and CIP were determined for the 27 NA-resistant *E. coli* isolates by using the broth microdilution method according to CLSI standards [7], with *E. coli* ATCC

Table 1. Prevalence of nalidixic acid (NA)- or ciprofloxacin (CIP)-resistant *Escherichia (E.) coli* isolates from anal samples

Sample sources	Number of anal samples	Number of <i>E. coli</i> isolates (%) [*]	Number of NA-resistant <i>E. coli</i> isolates (%) [†]	Number of CIP-resistant <i>E. coli</i> isolates (%) [†]
Companion animals	53	32 (60.4)	13 (40.6)	5 (15.6)
Pet-owners	17	9 (52.9)	3 (33.3)	1 (11.1)
Non-pet-owners	34	22 (64.7)	11 (50.0)	4 (18.2)
Total	104	63 (60.6)	27 (42.9)	10 (15.9)

^{*}The percentage value indicates the frequency of *E. coli* isolation from the anal samples. [†]The percentage value indicates the frequency of NA- or CIP-resistant isolates from the collected *E. coli* isolates. Note that all CIP-resistant isolates also showed resistance against NA.

25922 used as a reference strain.

Detection of PMQR genes and mutations in QRDRs

The 27 NA-resistant *E. coli* isolates were screened by PCR for the presence of the following PMQR genes: *aac(6′)-Ib-cr*, *qepA*, *qnrA*, *qnrB*, and *qnrS* [5,36,39]. The *cr* variant of the *aac(6′)-Ib* gene was identified by direct sequencing of the amplified *aac(6′)-Ib* gene [26]. Mutations in DNA gyrase (*gyrA* and *gyrB*) and topoisomerase IV (*parC* and *parE*) genes of the NA-resistant isolates were identified by using specific primers [2,10,32,37]. The wild-type *E. coli* K-12 sequence (GenBank accession No. U00096) was used as a reference [19].

Organic solvent tolerance (OST) assay

The activity of the AcrAB-TolC efflux pump system in NA-resistant *E. coli* isolates was measured by using the OST assay [35]. For efficiency-of-plating assays [35], cultures of isolates in logarithmic growth phase were diluted to an optical density of 0.2 at a wavelength of 530 nm, and 100-μL aliquots were spread onto Luria-Bertani (LB) agar, which was then overlaid with a mixture of hexane and cyclohexane [3:1 (v/v)]. The plates were sealed and incubated for 24 to 36 h at 30°C. The number of colonies was counted in triplicate and colony growth was recorded as confluent (++, ≥ 100 colonies), visible (+, < 100 colonies), or none (-). *E. coli* ATCC 25922 was used as a reference strain.

Evaluation of the effect of each (FQ)Q resistance mechanism on MICs of NA and CIP

To assess the relevance of the three (FQ)Q resistance

mechanisms to the increase in (FQ)Q resistance in *E. coli*, we evaluated the frequencies of point mutations in the QRDR region and PMQR genes as well as efflux pump activity in the 27 NA-resistant *E. coli* isolates. To assess the effects of point mutations and efflux pump activity, an additional comparison was made by determining Pearson’s correlation coefficient (*R*) [1]. The strength of efflux pump activity was graded based on the result of OST test described above (-, 0; +, 1; ++, 2). The *R* values were calculated between the number of point mutations or strength of efflux pump activity and MICs of NA or CIP by using SPSS software (ver. 23; IBM, USA) [1].

Results

Isolation of NA- or CIP-resistant *E. coli* from companion animals and humans

A total of 63 *E. coli* isolates (60.6%) were collected from 104 anal swab samples. Of these, 27 isolates (42.9%) were determined to be NA-resistant *E. coli*. Ten isolates (15.9%) were resistant to CIP, all of which also showed resistance to NA (Table 1). Overall, *E. coli* isolation and (FQ)Q resistance rates were higher in non-pet-owners than in pet-owners (Table 1).

Susceptibility of NA-resistant *E. coli* isolates to other antimicrobials

Of the 27 NA-resistant *E. coli* isolates, 23 (85.2%) were resistant to at least one additional antimicrobial (data not shown) and 20 (74.1%) were identified as MDR (Table 2). Antibiogram analysis revealed that more than half of NA-resistant isolates were also resistant to AM (17/27, 63.0%),

Table 2. Additional antimicrobial resistance profiling of 27 nalidixic acid (NA)-resistant *Escherichia (E.) coli* isolates

Source of NA-resistant <i>E. coli</i> isolates		AM	AMC	CAZ	CTT	GM	TE	CIP	SXT	C	ATM	CRO	CTX	MDR
Humans	Owners (n = 3)	1	1	0	0	1	1	1	1	0	0	0	0	2
	Non-owners (n = 11)	9	9	0	0	2	7	4	5	0	0	1	1	10
	Subtotal (n = 14)	10 (71.4)*	10 (71.4)	0 (0.0)	0 (0.0)	3 (21.4)	8 (57.1)	5 (35.7)	6 (42.9)	0 (0.0)	0 (0.0)	1 (7.1)	1 (7.1)	12 (85.7)
Companion animals	Dogs (n = 12)	7	7	3	4	3	6	5	3	2	2	3	3	8
	Cat (n = 1)	0	0	0	0	0	0	0	1	0	0	0	0	0
	Subtotal (n = 13)	7 (53.8)	7 (53.8)	3 (23.1)	4 (30.8)	3 (23.1)	6 (46.2)	5 (38.5)	4 (30.8)	2 (15.4)	2 (15.4)	3 (23.1)	3 (23.1)	8 (61.5)
Total	N = 27	17 (63.0)	17 (63.0)	3 (11.1)	4 (14.8)	6 (22.2)	14 (51.9)	10† (37.0)	10 (37.0)	2 (7.4)	2 (7.4)	4 (14.8)	4 (14.8)	20 (74.1)

*The frequency of *E. coli* isolates showing resistance to each used antimicrobial is shown in the parenthesis. †All 10 CIP-resistant isolates were determined as multi-drug resistances (MDRs). AM, ampicillin; AMC, amoxicillin/clavulanic acid; CAZ, ceftazidime; CTT, cefotetan; GM, gentamicin; TE, tetracycline; CIP, ciprofloxacin; SXT, sulfamethoxazole/trimethoprim; C, chloramphenicol; ATM, aztreonam; CRO, ceftriaxone; CTX, cefotaxime.

Table 3. Determination of the minimum inhibitory concentrations (MICs) and characterization of 27 nalidixic acid (NA)-resistant *Escherichia (E.) coli* isolates

Isolate Number	Source	NA MIC (µg/mL)	CIP MIC (µg/mL)	Mutations in the QRDRs*				Growth in presence of organic solvent [†]
				GyrA	ParC	GyrB	ParE	Mixture of hexane and cyclohexane [3:1 (vol/vol)]
P127	Owner	> 1,024	32	S83L D87N	S80I	— [‡]	L416F	—
P128	Owner	256	0.5	S83L	—	—	—	—
P143	Owner	256	0.25	S83L	—	—	—	+ +
P97	Non-owner	256	0.25	S83L	—	—	—	+
P98	Non-owner	128	0.25	S83L	—	—	—	—
P102	Non-owner	> 1,024	32	S83L D87N	S80I	—	L416F	—
P103	Non-owner	> 1,024	32	S83L D87N	S80I	—	L416F	—
P108	Non-owner	256	0.25	S83L	—	—	—	+ +
P109	Non-owner	512	1	S83L	—	—	—	+ +
P99	Non-owner	> 1,024	16	S83L D87N	S80I	—	L416F	—
P100	Non-owner	256	0.25	S83L	—	—	—	+ +
P154	Non-owner	> 1,024	16	S83L D87N	S80I	—	L416F	—
P144	Non-owner	128	0.25	S83L	—	—	—	+
P150	Non-owner	512	0.25	S83L	—	—	—	—
K132	Dog	256	0.125	S83L	—	—	—	+ +
R14	Dog	256	0.5	S83L	—	—	—	+ +
K161	Dog	> 1,024	256	S83L D87N	S80I	—	S458A	+ +
K48	Dog	> 1,024	64	S83L D87N	S80I E84G	—	I355T	+ +
K56	Dog	> 1,024	64	S83L D87N	S80I	—	S458A	+ +
K73	Dog	> 1,024	128	S83L D87N	S80I	—	S458A	+
K154	Dog	256	0.25	S83L	—	—	—	+ +
R56	Dog	128	0.125	D87N	—	—	—	+ +
K168	Dog	> 1,024	32	S83L D87N	S80I	—	S458A	+ +
J11	Dog	512	0.5	S83L	—	—	—	+
P129	Dog	256	0.25	S83L	—	—	—	+ +
K328	Dog	128	0.125	S83L	—	—	—	+ +
P123	Cat	256	0.25	S83L	—	—	L477M	—
ATCC 25922	Control	8	0.008	—	—	—	—	—

*The identified codon sites of mutations in NA-resistant *E. coli* isolates are indicated. [†]Scoring: + +, confluent growth (≥ 100 colonies); +, visible growth (< 100 colonies); —, no growth. [‡]No mutations were found in the target gene. CIP, ciprofloxacin; QRDRs, Q resistance-determining regions.

AMC (17/27, 63.0%), and TE (14/27, 51.9%). Ten of the NA-resistant isolates showed resistance to CIP (10/27, 37.0%); interestingly, these were all identified as MDR (Table 2). On the contrary, resistances against CAZ, CTT, C, ATM, CRO, and CTX were relatively low. In addition, 3 of the 27 NA-resistant isolates (11.1%) were identified as ESBL-producing *E. coli* harboring the *bla*_{CTX-M} gene and were obtained from two dogs and a non-pet-owner (data not shown).

Determination of MICs

The MICs of NA for the 27 NA-resistant *E. coli* isolates ranged from 128 to > 1024 µg/mL. Ten isolates showed much higher MICs of NA (> 1024 µg/mL) than the others, and only those isolates showed resistance to CIP (MICs of CIP, 16–256 µg/mL). The remaining isolates showed low MICs of CIP, ranging from 0.125 to 1 µg/mL (Table 3).

Analysis of mutations in QRDRs and detection of PMQR genes

Mutations were identified as nucleotide alterations in QRDRs responsible for changes in amino acid sequences in the protein products. Mutations were detected in the *gyrA* gene in all NA-resistant isolates (Table 3); 10/27 (37.0%) had double amino acid substitutions (S83L and D87N); 16/27 (59.3%) had a single amino acid substitution (S83L); and 1/27 (3.7%) had a single D87N substitution. Mutations in the *parC* gene were detected in 10/27 isolates (37.0%); nine (33.3%) had a single S80I mutation, and one had double mutations of S80I and E84G. However, no mutations were found in the *gyrB* gene. Eleven isolates (40.7%) had mutations in codons 355, 416, 458, or 477 of the *parE* gene (Table 3).

Notably, all 10 isolates with double amino acid substitutions in GyrA had point mutations in both *parC* and *parE* genes, and only these isolates were resistant to CIP. While *parC* mutations were detected only in these 10 isolates, *parE* mutations were found in those 10 isolates and an additional isolate with a single amino acid substitution in the *gyrA* gene (No. P123; Table 3). Of the 10 CIP-resistant isolates, five originating from humans had an amino acid substitution in only codon 416 of *parE* (L416F). In contrast, the other five CIP-resistant *E. coli* isolates from dogs had amino acid substitutions in codon 355 or 458 of *parE* (I355T or S458A) (Table 3). A PMQR gene was detected in only one isolate (No. K73), which harbored the *aac*(6′)-*Ib-cr* gene encoding a CIP-modifying enzyme (data not shown).

Measurement of efflux pump activity

Efflux pump activity in the 27 NA-resistant *E. coli* isolates was measured by using the OST assay as described in the Materials and Methods section. A total of 14 (51.9%) and 4 (14.8%) isolates showed confluent and visible growth, respectively, on the organic solvent mixture (Table 3).

Relative contribution of each (F)QQR resistance mechanism to increases in MIC

We analyzed the correlations between efflux pump activity or target mutations and MICs of NA or CIP in the 27 NA-resistant isolates and observed that the MICs of both NA and CIP were highly correlated with the number of point mutations in the QRDR ($R = 0.878$ and 0.954 , respectively; Fig. 1). However, efflux pump activity was not correlated with either the NA MIC ($R = -0.239$) or the CIP MIC ($R = -0.169$) in NA-resistant *E. coli* isolates (Fig. 1).

Discussion

Many epidemiological studies have reported that companion animals in households are potential sources of transmissible bacteria such as *E. coli* and *Salmonella* spp. (reviewed in [9]). However, in the present study, the isolation and antimicrobial resistance rates of *E. coli* were not higher in pet-owners than in non-pet-owners. Another study reported a lower risk of MDR staphylococci carriage in nursing home residents living with companion animals than in those living without pets [12]. These findings may suggest that although companion animals can be sources of bacterial infections in households, they do not always negatively affect the hygienic status of their owners.

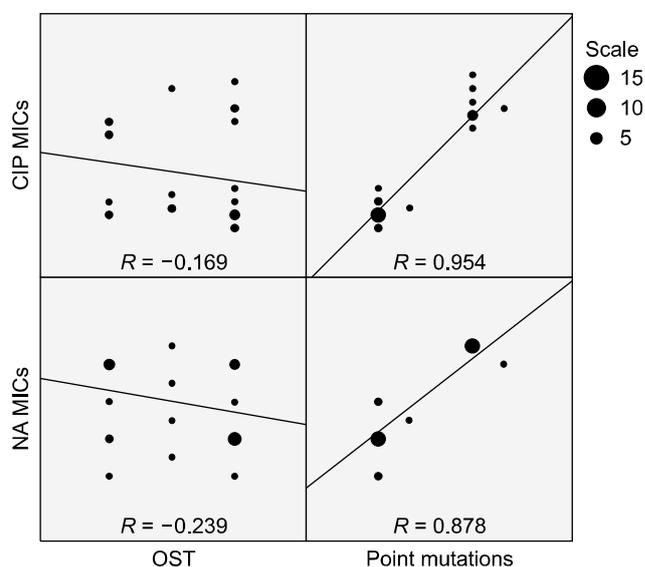


Fig. 1. Correlations between organic solvent tolerance (OST) or number of target mutations and minimum inhibitory concentrations (MICs) of nalidixic acid (NA) and ciprofloxacin (CIP) among 27 NA-resistant *E. coli* isolates. The size of the closed circle in each dot plot represents the number of NA- or CIP-resistant *E. coli* isolates. The scale box located on the right side of graphs shows three different-sized closed circles with the corresponding number of NA- or CIP-resistant isolates. The gradient of the trend line in each dot plot represents positive or negative correlation between the two variables. R : correlation coefficient.

(FQ)Q resistance is closely associated with MDR in *E. coli* [31]. We observed that about 74% of NA-resistant *E. coli* isolates were MDR. Cross-antimicrobial resistance between Q and β -lactams frequently occurs in *E. coli* and *Klebsiella* spp. due to the extensive use of antimicrobials against these bacteria in human and veterinary medicine [11]. The mechanism underlying the association between MDR and Q resistance is currently unclear. In our study, the MDR rate was higher in CIP-resistant than in NA-resistant *E. coli* isolates (100% vs. 74.1%). As shown in previous studies [13,15,19], as well as in the current study, CIP resistance is strongly correlated with multiple mutations in QRDRs.

In this study, all 27 NA-resistant *E. coli* had at least one target mutation in the QRDRs. Low or high efflux pump activity was observed in 18 of those isolates (66.7%), but only one (3.7%) harbored a PMQR gene. Mobile (FQ)Q resistance gene transfer by plasmids has been demonstrated, and high detection rates for PMQR genes in (FQ)Q-resistant *E. coli* isolated from human and animal specimens have been reported [23,24]. However, other studies have reported low detection rates or an absence of PMQR genes in FQ-resistant *E. coli* from animals [13,19]. The results of the present study are consistent with the latter and highlight the low prevalence of PMQR in Korea. The presence of PMQR genes is closely associated with that of genes encoding ESBL in *E. coli* isolates [18]. Accordingly, the isolate harboring *aac*-(6')-Ib-cr (No. K73) was determined to be an ESBL-producing *E. coli* strain carrying the *bla*_{CTX-M} gene in this study.

We found no correlation between increased efflux pump activity and increases in MICs of both NA and CIP in NA-resistant *E. coli*. When MICs in 15 isolates harboring the same single amino acid substitution in GyrA (S83L) were compared, to exclude other mutational variables in QRDRs, MICs in isolates with high OST were not higher than those in isolates with no OST (average MICs: NA, 277 vs. 299 μ g/mL and CIP, 0.33 vs. 0.33 μ g/mL). Efflux pumps reduce the concentration of substrates within cells via active transport, and an increase in their activity has been reported to contribute to reduced (FQ)Q susceptibility [27]. However, our results indicate that efflux pump activity did not contribute to resistance against (FQ)Q in these resistant isolates. If mutations in QRDRs lead to sufficiently high levels of (FQ)Q resistance in *E. coli*, the activity of the efflux pump may not further increase the MICs of (FQ)Q [1]. Similar findings were reported in other Gram-negative bacteria such as *Klebsiella pneumoniae* and *Campylobacter* spp. [8,29]. In contrast, the number of mutations in QRDRs was strongly correlated with increases in the MICs of both NA and CIP. Particularly, in QRDRs the (FQ)Q resistance in clinical *E. coli* isolates is more closely associated with mutations in the *gyrA* gene, whereas mutations in *gyrB*, *parC*, and *parE* genes are less important in the establishment of (FQ)Q resistance [16]. A single mutation in the

gyrA gene has been linked to low FQ resistance in *E. coli* [38], while high FQ resistance was reported to be acquired via accumulation of mutations in QRDRs [15]. In particular, FQ resistance was related to double amino acid substitutions in GyrA with or without mutations in *parC* and *parE* genes [13,19,20]. In the present study, all CIP-resistant isolates from both humans and animals had double point mutations in *gyrA* concurrent with target mutations in *parC* and *parE* genes. These results indicate that (FQ)Q resistance mechanisms in *E. coli* at animal hospitals are similar to those observed in humans.

Mutations in the *parC* or *parE* gene have been reported to be closely related to secondary mutations in the *gyrA* gene [4,20]. Consistent with those results, all mutations in *parC* or *parE* were detected in isolates with double amino acid substitutions, except for one case. Among the four amino acid substitutions in ParE identified in this study, three have been previously reported (I355T, L416F, and S458A) [19,25]. However, the amino acid substitution (L477M) found in a cat isolate (No. P123) is a novel finding. Interestingly, the patterns of *parE* mutations observed in the NA-resistant *E. coli* isolates were distinct in each species; L477M and L416F were present only in cat and human isolates, respectively. All I355T and S458A substitutions were only found in dog isolates, and they were in conjunction with amino acid substitutions in both GyrA and ParC. Taken together, the CIP-resistant isolates from humans were distinct from those obtained from dogs and cats in terms of the position of the *parE* gene mutations.

In conclusion, this is the first study to examine the mechanisms of (FQ)Q resistance in NA-resistant *E. coli* isolates from companion animals and their owners compared to those from non-pet-owners. The rates of *E. coli* isolation and (FQ)Q resistance were not higher in pet-owners than in non-pet-owners, which may suggest that persons living with pets are not always at a higher risk of bacterial infection than those living without pets. The prevalence of PMQR genes was very low and efflux pump activity was not found to contribute alone to the acquisition of high-level (FQ)Q resistance. Target site alterations in QRDRs appeared to be the most important mechanism contributing to high-level (FQ)Q resistance in *E. coli* of both animal and human origins in Korea. Since the (FQ)Q resistance mechanisms in pet isolates were the same as those found in human isolates, prudent use of (FQ)Q by veterinarians is warranted in order to prevent the development and dissemination of (FQ)Q-resistant bacteria. To this end, a continuous monitoring process in veterinary hospitals may be needed to identify the trends and dissemination of (FQ)Q resistance in companion animals and their owners.

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Conflict of Interest

The authors declare no conflicts of interest.

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